

1 **Systemic sodium selenate, a PP2A activator, attenuates con-**
2 **ditioned taste aversion learning and phospho-MAP kinase in-**
3 **duction**

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12 cleus

14 **Abstract**

15 [REVISE, especially to include c-FOS]

16 CTA learning is constrained by phosphatase activity and is enhanced by, e.g., okadaic acid
17 (Oberbeck 2010) or calcineurin knockdown (Baumgartel 2008). Conversely, phosphatase activation
18 should attenuate CTA. Recently, sodium selenate was found to be a specific activator of protein
19 phosphatase 2A (PP2A; Corcoran 2010, van Eersel 2010). To characterize the effects of selenate
20 on CTA, we determined 1) the dose of selenate which induces a CTA and 2) if a subthreshold dose
21 of selenate would attenuate a LiCl-induced CTA. Water-restricted rats were given 10-min access
22 to 0.125% saccharin, then injected with selenate (0, 0.5, 1, or 2 mg/kg ip, n=6 /dose). The next
23 day, 2-bottle 24-h preference extinction tests of water vs. saccharin were begun and continued for
24 14 days. Rats that received saccharin paired with 1 or 2 mg/kg selenate acquired a CTA with
25 reduced saccharin preference across extinction, but rats receiving 0 or 0.5 mg/kg showed a high
26 preference and thus no CTA. A separate group of rats were injected on conditioning day with the
27 low dose of selenate (0.5 mg/kg) or saline. 2h later, the rats were given 10min access to saccharin,
28 then injected with either NaCl or LiCl (0.15 M, 6 ml/kg ip). Preference tests were run for 14 days.
29 Vehicle-LiCl rats showed a persistent decrease in saccharin preference. While selenate-pretreated
30 rats showed reduced saccharin preferences, their preferences were higher than vehicle-LiCl rats. This
31 suggests that selenate interfered with the rats' ability to acquire a CTA, consistent with phosphatase
32 activation.

1 Introduction

The reversible phosphorylation of specific intracellular substrates in several brain regions has been shown necessary for learning and memory formation in many learning paradigms (1) (2) (3). Protein kinases and phosphatases are responsible for the dynamic regulation of phosphorylation states of signaling protein substrates, and ultimately the cellular events that correlate with learning and memory, such as synaptogenesis and synaptic plasticity (4) (5). In the past, more research has focused on the role of kinases and positive regulation of substrate phosphorylation in learning and memory; however, recent interest has risen in the role of phosphatases and negative regulation of substrate phosphorylation.

Protein phosphatase 2A (PP2A) is a serine/threonine phosphatase that, in combination with PP1 and PP2B (calcineurin), is responsible for over 90% of the phosphatase activity in neurons (6). PP2A has a heterotrimeric arrangement in which the A subunit serves as a scaffolding subunit, the B subunit serves as a regulatory subunit, and the C subunit serves as a catalytic subunit. The pleiotropic effects of PP2A are regulated spatially and temporally by the attachment of the A subunit, various B subunits, and endogenous inhibitors. Although PP2A has several endogenous modulators, there are not many viable experimental tools for studying phosphatase dependent processes of learning and memory in vivo. As a result, it is important to find new chemicals with increased specificity for phosphatases to better define their activity (2).

1.1 Sodium Selenate and PP2A Activation

Recently, Corcoran et al. found that sodium selenate (Na_2SeO_4) is a specific and potent activator of the PP2A heterotrimer (7). Several forms of selenium have been tested for possible pharmacological applications in models of Alzheimer's disease, in which enhancement of PP2A activity leads to a decrease in hyperphosphorylated tau protein (7)(8). A possible mechanism for selenate activation of PP2A is increased expression of the PR55 beta subunit, an obligate activator of PP2A for tau dephosphorylation (9).

58 1.2 [details of selenate and other forms of Learning Acquisition]

59 Traumatic brain injury in rats leads to increased phospho-tau and decreased PP2A activity; chronic
60 selenate treatment (1 mg/kg/day for 12 weeks via osmotic mini-pump) improved behavioral deficits
61 (such as water maze performance) increased PP2A activity , and decreased levels of tau phospho-
62 rylation(9).

63 Reduced tau phosphorylation in human neuroblastoma cells. In rats, acute sodium selenate
64 (10-35 mg/kg, i.p) administered 2 h before stimulation reduced 6-Hz electrical stimulation seizures;
65 chronic access to selenate in drinking water (0.12 or 1.2 mg/100 ml = 0.64 - 6.4 micromolar) reduced
66 seizures induced by pentylenetetrazol (PTZ) or amygdalar stimulation (10).

67 In a brain slice preparation, selenate rescued deficits in long-term depression in THY-Tau22
68 mice which display tau hyperphosphorylation and impaired memory, as well as reversing the effects
69 of okadaic acid (11).

70 1.3 Selenium Toxicity

71 Although selenium is an essential micronutrient, and sodium selenate appears to have superior
72 characteristics for in vivo applications related to learning and memory, selenium is also toxic at
73 doses not much higher than the dietary requirement (12)(13). The mammalian selenoproteome
74 consists of some 2 dozen selenocysteine-containing proteins (24 in rodents, 25 in human) including the
75 glutathione peroxidase, thioredoxin reductase and iodothyronine deiodinase enzymes (14) . Selenium
76 toxicity is likely due to the generation of superoxide and other reactive oxygen species, the oxidation
77 of thiols, and by substitution for sulfur in methionine to form selenomethione which then may be
78 incorporated into many sulfur-containing proteins (15) .

79 Aversive effects of selenium compounds, especially at low sublethal doses, is by observing
80 reduced intake, due to acute toxicity, and by learned food or fluid avoidance after conditioned taste
81 aversion (CTA) acquisition. In CTA learning, animals will reduce subsequent intake of a diet if

82 consumption was previously paired with adverse consequences, even if the toxic effects were delayed
83 for hours after consumption.

84 Reduced intake and avoidance of toxic diets containing selenium compounds has been demon-
85 strated in many species. Because selenium is concentrated by certain species of plants in selenium-
86 rich geographies, selenosis is a potential hazard for livestock and native herbivores. Waste from
87 mining or agricultural run-off can also introduce selenium into the environment and hence into the
88 food-chain. It has therefore been of interest to determine whether species can avoid selenium-rich
89 food sources due to an aversive taste or odor of selenium compounds, or acute toxic effects, or
90 learned food aversions. Aphids (16) (17), crickets and grasshoppers (18) , southern armyworms (19)
91 , mallard ducks (20) , owls (21), prairie dogs (22) , sheep (12) (23), pigs (24), and cattle (12) have
92 been shown to reduce intake of selenium-containing foods, or show reduced preference for selenium-
93 containing foods over control diets. However, it has also been observed that honey bee foragers do
94 not reduce their reflexive ingestive responses nor their consumption of selenium-adulterated sucrose
95 solutions, even at toxic concentrations(25). Similarly, Argentine ants do not show decreased pref-
96 erence for sucrose containing selenium compounds over plain sucrose in short-term choice tests (26).
97 Thus, the ability to detect or tolerate dietary selenium varies across species.

98 In laboratory studies, rats show a preference for control or low selenium-diets over high
99 selenium diets (27) and [Franke and Potter 1935]. Consistent with the effects of other selenium
100 compounds, selenate adulteration also causes a reduction of food (28)(29) (30) or water intake (31)
101 (32) Acute injection of selenate can also reduce food intake and body weight gain (33)

102 Only a few of these studies (e.g. (12)(23) (20)) have determined if animals were responding
103 to an acute perception of selenium compounds, or if the animals used food-associated cues, to avoid
104 consumption, after acquiring a selenium-induced conditioned food aversion. Selenium absorbing
105 plants may themselves have salient taste or odor: for example, Provenza et al. have demonstrated
106 that the odor of *Astragalus bisulcatus*, a sulfur-containing selenium-absorbing plant, can serve as
107 a CS that reduced preference for CS+barley-straw after pairing with oral-intubation of LiCl (34).
108 Selenium compounds may also present distinctive cues. In particular, dimethyldiselenide, a major
109 and volatile metabolite of selenite and selenate, has a pungent garlic odor to humans, that may be
110 innately aversive or serve as an indicative CS in learned selenium aversions.

111 1.4 Conditioned Taste Aversion

112 Human phase 1, dose escalation study from 5 to 90 mg / day (tds) in patients with prostate cancer:
113 dose limiting toxicity at 90 mg from fatigue, or diarrhea and muscle spasms (35). also reports of
114 nausea and vomiting.

115 12 ug/ml for 12 weeks -> no effect on fluid intake in mice; also showed that selenate treated
116 mice show preference for saccharin same as untreated mice (but not a CTA test, just unconditioned
117 taste detection test). (8) .

118 Previous studies have found that when selenate (2.5-3.8 ug/ml) is added to their drinking
119 water, rats reduce their fluid intake by 20-25% (31). One of the more recent studies found that
120 administering

121 1.5 Goals

122 However, Na_2SeO_4 has not been utilized in CTA experiments, nor has a dose-response curve been
123 characterized for its toxicity. The following experiments were designed to elucidate the dose-response
124 relationship between systemic administration of Na_2SeO_4 and its potential aversive effects as mea-
125 sured by CTA induction and c-Fos induction in the visceral neuraxis. , and to determine the effect
126 of Na_2SeO_4 administration on LiCl-induced CTA.

127 The goals of this study were

128 i. to determine a range of doses at which systemic sodium selenate paired with saccharin
129 caused in CTA acquisition and activated the visceral neuraxis as measured by c-Fos induction, and

130 ii. to assess the ability of a low dose of sodium selenate to attenuate acquisition of a
131 LiCl-induced CTA, and to attenuate neural activation as measured by reductions in c-Fos and

132 phospho-MAP kinase levels.

133 2 Methods

134 2.1 Animals

135 Adult male Sprague-Dawley rats (Charles River) were housed individually in polycarbonate cages
136 in a temperature-controlled (22 ± 2 °C, 30-40% humidity) colony room. The rats were maintained
137 on a 12 h light/dark cycle with lights-on at 7:00 A.M. All procedures were conducted during the
138 light cycle. The rats had ad libitum access to Purina Rodent Chow and deionized-distilled water
139 except as noted. All procedures were approved by the Florida State University animal care and use
140 committee.

141 2.2 Experiment 1. To Investigate the Dose-Response Relationship for 142 Sodium Selenate as the Unconditional Stimulus (US) in CTA Learn- 143 ing.

144 Na_2SeO_4 is a known toxin at excessive doses. In order to determine the toxicity of Na_2SeO_4 , I
145 measured rats' CTA acquisition after a variable dose of Na_2SeO_4 was paired with saccharin. Rats
146 received access to the CS (saccharin) for 10 minutes and then received 1 of 4 doses of the US (0,
147 0.5, 1 or 2 mg/kg Na_2SeO_4 i.p.). A 2 bottle 24-hour preference test was used to measure CTA. It
148 was predicted that rats given a higher dose (e.g. 1 and 2 mg/kg) of Na_2SeO_4 would acquire a CTA to
149 saccharin; while those that received a low dose (0.5 mg/kg) and the vehicle would not.

150 Adult male Sprague-Dawley rats (n=24) were water restricted for 8 days. During this
151 restriction, the rats received access to water for a period of time that was reduced every other day
152 until rats had only 10 min water access a day. This ensured that the rats would sample the CS
153 provided to them on the conditioning day. The next day, rats were given 10-min access to the CS
154 (0.125% saccharin). Saccharin bottles were then measured in order to ensure that every rat sampled
155 the CS. If any rat had not sampled the US, the bottle was placed on the cage for an additional 5
156 min. Twenty minutes later, the rats were injected with Na_2SeO_4 at 4 doses (0, 0.5, 1, or 2 mg/kg,

157 n = 6/dose); which served as the US. The next day rats underwent 2-bottle 24-h preference tests
158 for 14 days. This preference test is comprised of giving the rats access to 2 bottles, one filled with
159 water and one filled with the CS (0.125% saccharin), and weighing each bottle daily. Preference
160 was calculated as (saccharin intake) / (total intake). Bottle location was switched daily in order to
161 observe any place preference that could have confounded the association.

162 **2.3 Experiment 2. To Investigate the Level of c-Fos Expression in the** 163 **Visceral Neuraxis after Administration of Various Doses of Sodium** 164 **Selenate.**

165 In order to determine if Na₂SeO₄ elicits the activation of brain areas associated with the visceral
166 neuraxis in a dose dependent manner, I injected rats with variable doses of Na₂SeO₄ and measured
167 brain activation via c-Fos immunoreactivity. Rats were injected with 0, 0.5, 1, 2 mg/kg Na₂SeO₄
168 or 12 ml/kg of 0.15 M LiCl, euthanized, and then perfused. Region specific brain activation was
169 accessed by measuring levels of c-Fos expression via DAB immunohistochemistry. It was predicted
170 that levels of c-Fos expression in the visceral neuraxis would be higher in rats injected with high
171 doses of Na₂SeO₄ (1 and 2 mg/kg) and c-Fos expression would be lower in rats injected with 0.5
172 mg/kg Na₂SeO₄ or the vehicle.

173 Adult male Sprague-Dawley rats (n=30) were injected with doses of Na₂SeO₄ comparable
174 to those used in experiment 1 (0, 0.5, 1, or 2 mg/kg Na₂SeO₄) or a 12 ml/kg dose of LiCl (0.15 M)
175 to serve as a positive control. 2 hours later, rats were euthanized via i.p. injection of Nembutal and
176 then perfused transcardially with 100 mL preperfusion solution and then 400mL paraformaldehyde.
177 Brains were left to postfix for 2 hours and then stored in 30% sucrose. Forebrain and hindbrain
178 sections of 40 micrometers were cut via microtome. Sections were then processed with goat anti-c-
179 Fos antisera (Santa Cruz) using DAB chromogenic immunohistochemistry. For c-Fos quantification,
180 the cells expressing darkly positive, nuclear staining were quantified with custom software (MindsEye,
181 T. Houpt; Kwon et al., 2008). c-Fos was quantified in regions of the brain associated with CTA and
182 the visceral neuraxis (i.e. central amygdala (CeA), paraventricular nucleus (PVN), nucleus of the
183 solitary tract (NTS), parabrachial nucleus (PBN), and supraoptic nucleus (SON)).

2.4 Experiment 3. To Investigate whether Activation of PP2A via a Low Dose of Sodium Selenate will Attenuate a LiCl-Induced CTA.

In order to determine whether PP2A interferes with CTA acquisition, I pretreated rats with a nontoxic dose of Na_2SeO_4 (a selective PP2A agonist) or vehicle and then measured CTA acquisition. Rats were initially separated into two groups and then received an i.p. injection of Na_2SeO_4 or vehicle. In order to elicit a CTA, rats in each group then received access to a CS (saccharin) which was paired with a US (6 ml/kg, 0.15 M LiCl) or vehicle. In order to test Na_2SeO_4 's effects on a learned CTA, rats were then subjected to a 14 day 2-bottle 24-h preference test. It was predicted that the administration of an appropriate dose of sodium selenate would result in an attenuation of CTA learning via PP2A activation.

Adult male Sprague-Dawley rats ($n=24$) were water restricted for 8 days. The duration rats had access to water was reduced throughout the 8 days until rats were limited to 10 min water access a day. This ensured that the rats would sample the CS presented on the conditioning day. On the day of conditioning, rats were injected with either the low dose of Na_2SeO_4 (0.5 mg/kg) or a saline vehicle. This dose of Na_2SeO_4 was used because it did not elicit a CTA in experiment 1; therefore it was concluded to be a safe dose to study Na_2SeO_4 's effects on learning and memory processes without the risk of resulting in an exaggerated US. Two hours later, the rats were given 10-min access to the CS (0.125% saccharin). Saccharin bottles were then measured in order to ensure every rat sampled the CS. If any rat did not sample the US, the bottle was placed on the cage for an additional 5 min and weighed again. Twenty minutes later, the rats received i.p. injections of either the US (6 ml/kg, 0.15 M LiCl) or vehicle (NaCl). This separated the rats into 4 groups: vehicle-NaCl, vehicle-LiCl, Na_2SeO_4 -NaCl, and Na_2SeO_4 -LiCl ($n=6$ /group). The next day rats underwent 2 bottle 24-h preference tests that continued for 14 days. The preference test is comprised of allowing the rats access to 2 bottles, one filled with water and one filled with the CS (0.125% saccharin), and then recording the weight of each bottle daily. Preference was calculated as (saccharin intake) / (total intake). The bottle location of each tastant was switched daily in order to observe any place preference that may have confounded the association.

2.5 Experiment 4. Sodium Selenate Attenuation of c-Fos and phospho-MAP kinase induction

To assess the effects of Na₂SeO₄ pretreatment on neuronal activation in the visceral neuraxis, c-Fos and phospho-MAP kinase (pMAPK) induced by LiCl was examined with and without Na₂SeO₄ pretreatment

Adult male Sprague-Dawley rats (n=???) were injected with either Na₂SeO₄ (0.5 mg/kg) or vehicle (1 ml/kg 0.15 M NaCl). One hour later, rats were injected with either LiCl or NaCl (0.15M, 12 mg/kg), counterbalanced across selenate and vehicle groups. At 1 hour after the second injection, rats were euthanized via i.p. injection of Nembutal and then perfused transcardially with 100 mL preperfusion solution and then 400mL paraformaldehyde.

Brains were left to postfix for 2 hours and then stored in 30% sucrose. Forebrain and hindbrain sections of 40 micrometers were cut via microtome. Alternate sections were processed with goat anti-c-Fos antisera (Ab#, Santa Cruz) and anti-pMAPK (Ab#, Cell Signaling) using DAB chromogenic immunohistochemistry. c-Fos positive nuclei and pMAPK positive soma were quantified as in Experiment 2.

2.6 Statistical Analysis

Data are presented as the mean \pm standard error of the mean. Preference scores were analyzed using Statistica software (Statsoft, Tulsa, OK), using either one-way or two-way, repeated measures analysis of variance (ANOVAs). For analysis of CTA magnitude, Treatment group (0.0, 0.5, 1, or 2 mg/kg Na₂SeO₄) served as the independent variable and saccharin preference and the first day of 2-bottle preference testing, served as the dependent variable for the one-way ANOVA. For analysis of CTA Extinction in Experiments 1 and 3, Treatment group (Experiment 1: 0.0, 0.5, 1, or 2 mg/kg Na₂SeO₄; Experiment 3: vehicle-vehicle, vehicle-LiCl, selenate-vehicle, and selenate-LiCl) served as the first independent variable, day of testing served as the second independent variable, and saccharin preference served as the dependent variable for the two-way repeated measures ANOVAs.

236 For analysis of c-Fos induction in Experiment 2, Treatment group (0.0, 0.5, 1, 2 mg/kg
237 Na₂SeO₄, or 76 mg/kg LiCl) served as the independent variable and c-Fos counts in each brain
238 region served as the dependent variable for a one-way ANOVA. Tukey-Kramer Highly Significant
239 Difference pairwise comparisons were used for all post-hoc analyses.

240 3 Results

241 3.1 Experiment 1

242 A one-way, ANOVA revealed a significant main effect of treatment on the first day of 2-bottle
243 preference tests ($F(3, 20) = 13.28, p = 0.000054$) (see figure 1). Post hoc analysis revealed that
244 the mean saccharin preference of the 1 mg/kg selenate treatment group and the 2 mg/kg selenate
245 treatment group were significantly lower than the mean of the 0.0 mg/kg selenate treatment group
246 (negative control). The mean saccharin preference of the 2 mg/kg selenate treatment group was
247 significantly lower than the 0.5 mg/kg selenate treatment group.

248 A two-way, repeated measures ANOVA across the 14 days of extinction testing revealed a
249 significant main effect of the treatment ($F(3,39) = 4.45, p = 0.015$) and for day of testing ($F(13, 39)$
250 $= 4.82, p = 0.0000$), but no interaction between the two factors (see figure 2). Post hoc analysis
251 showed that the mean saccharin preference for the 0.5 mg/kg selenate treatment group was not
252 significantly different from 0.0 mg/kg selenate treatment group (negative control group) across all
253 14 days of the 2-bottle preference tests. However, the mean saccharin preference for the 1 mg/kg
254 selenate treatment group was significantly lower than the 0 mg/kg selenate control group on days
255 1-5 and 7-9 during the 14 days of 2-bottle preference testing. The mean saccharin preference for the
256 2 mg/kg selenate treatment group was significantly lower than the control group on days 1-13 of
257 the 2-bottle preference test, but was not lower on day 14. Therefore, it was concluded that the high
258 doses of sodium selenate (1mg/kg and 2 mg/kg doses) can serve as a CS if paired with saccharin and
259 induce a conditioned taste aversion without the addition of another toxin; however, the low dose
260 of sodium selenate (0.5 mg/kg dose) does not serve as a US when paired with saccharin and is not
261 sufficient to induce a CTA.

262 3.2 Experiment 2

263 One-way ANOVAs revealed that injections of sodium selenate and/or LiCl significantly induced c-
264 Fos in the NTS ($F(4,20) = 2.90, p < 0.05$), PBN ($F(4,22) = 3.67, p < 0.05$), CeA ($F(4,25) = 3.63,$
265 $p < 0.05$), PVN ($F(4,25) = 6.54, p < 0.001$), and SON ($F(4,24) = 4.55, p < 0.01$) (see figures 3, 4,
266 5, 6 & 7).

267 In the NTS, despite a significant overall effect of treatment, post hoc analyses showed no
268 significant differences between any of the treatment groups and either of the control groups (negative
269 or positive; see figure 3). Furthermore, in the NTS, none of the groups were significantly different
270 from each other.

271 In the PBN, both the LiCl treatment group (positive control group) and the 2 mg/kg
272 selenate treatment group induced significantly higher c-Fos counts than the vehicle (negative control
273 group) (see figure 4). No other treatment group was significantly different from either control group.

274 In the CeA, only the 1mg/kg selenate treatment group showed a significantly higher c-
275 Fos count than the vehicle; although elevated, LiCl-induced c-Fos counts failed to reach statistical
276 significance (see figure 5). In the PVN, the LiCl injection induced significantly higher c-Fos than
277 the vehicle; however, all other doses of selenate failed to induce significantly more c-Fos than the
278 vehicle (see figure 6).

279 In the SON, selenate injections failed to induce any c-Fos and only the LiCl injections
280 induced significantly more c-Fos than the vehicle (see figure 7). Therefore, it was concluded that,
281 despite sodium selenate's lack of a uniform effect on c-Fos induction across all of the brain areas
282 that were observed, sodium selenate induced c-Fos in patterns specific to neuroanatomical functional
283 groups. In the brainstem (NTS and PBN), sodium selenate induced c-Fos in a dose dependent
284 manner and this pattern possibly occurred in the CeA as well. However, in the hypothalamus (SON
285 and PVN), none of the doses of sodium selenate induced c-Fos.

286 3.3 Experiment 3

287 A one-way ANOVA revealed a significant main effect of treatment on the first day of the 2-bottle
288 preference test ($F(1, 20) = 20.83, p = 0.0002$) (see figure 8). Post hoc analysis showed that the
289 mean saccharin preference of the NaCl-LiCl treatment group (positive control) was significantly
290 lower than the mean saccharin preference of the NaCl-NaCl treatment group (negative control) on
291 the first day of 2-bottle preference tests. The mean saccharin preference of the NaCl-LiCl treatment
292 group (positive control) was significantly lower than the mean saccharin preference of the selenate-
293 NaCl treatment group on the first day of 2-bottle preference tests. The selenate-LiCl group was
294 not significantly different from the NaCl-NaCl treatment group (negative control) on the first day
295 of 2-bottle preference tests, but had a p-value ($p = 0.0628$) that was very close to significance.

296 Two-way, repeated measures ANOVA across the 14 day extinction testing revealed a sig-
297 nificant main effect of the treatment ($F(3,39) = 4.56, p = 0.014$) and of day of testing ($F(13, 39)$
298 $= 6.80, p = 0.0000$), but no interaction between the two factors (see figure 9). Post hoc analysis
299 revealed that the mean preference of the NaCl-LiCl treatment group (positive control group) was
300 significantly lower than the NaCl-NaCl treatment group (negative control group) on all 14 days
301 of the 2-bottle preference test. The mean preference for selenate-NaCl treatment group did not
302 show a significant difference from the negative control group; however the mean preference of the
303 selenate-NaCl treatment group was significantly higher than the positive controls for the full 14
304 days of the 2-bottle preference test. The mean preference for the selenate-LiCl treatment group
305 was significantly lower than the NaCl-NaCl and selenate-NaCl groups on the first day, but failed
306 to show a statistical difference on days 2-14 of the 2-bottle preference test. Furthermore, the mean
307 preference of the selenate-LiCl group was significantly higher than the NaCl-LiCl group on days
308 5-9 of the 2-bottle preference test. Therefore, it was concluded that a low dose of sodium selenate
309 (0.5 mg/kg) interferes with the learning and memory processes in conditioned taste aversion and
310 increases the rate of memory extinction for the conditioned taste aversion.

311 3.4 Experiment 4

312 4 Discussion

313 Though it has been known for some time that phosphorylation of signaling protein substrates plays
314 an important role in learning and memory, much of the research delineating the role of this particu-
315 lar intracellular signaling mechanism has focused specifically on the role of kinases. However, recent
316 interest has risen in the role of phosphatases and negative regulation of signaling protein substrate
317 phosphorylation in learning and memory. In 2010, Oberbeck et al. found that direct administra-
318 tion of okadaic acid, a known inhibitor of PP1 and PP2A, to the amygdala enhances learning of
319 conditioned taste aversions and makes the memory of these aversions resistant to extinction. While
320 studies looking at the inhibition of phosphatases are useful, they are incomplete alone. It is also im-
321 portant to look at the behavioral and intracellular effects of phosphatase activation and to increase
322 the specificity of the pharmacological agents modulating phosphatase activities.

323 Corcoran et. al (2010) proposed that sodium selenate is a potent and specific activator
324 of the PP2A heterotrimer, which makes Na_2SeO_4 an excellent candidate drug to study the role
325 of PP2A on learning and memory. Selenium is an essential micronutrient that is incorporated in
326 selenoproteins required for antioxidant defense, thyroid hormone production, and immune responses.
327 However, Na_2SeO_4 is known to have toxic effects at doses that are only marginally higher than what
328 is required by diet (Gasmi , 1997; Stowe, 1992). The proximal cause of selenium toxicity is unknown;
329 however selenosis can result in cirrhosis of the liver, pulmonary edema, and death (Agency for Toxic
330 Substances and Disease Registry, 2003). Nevertheless, Na_2SeO_4 is the best drug candidate out of all
331 of the forms of selenium salt because it is the least toxic form of selenium, it shows no neurotoxicity,
332 and it is the only form that activates PP2A (Corcoran e.t al, 2010). In this thesis, I showed that
333 0.5 mg/kg Na_2SeO_4 does not have toxic effects that can induce a CTA, that Na_2SeO_4 activates the
334 visceral neuraxis and the CeA, and that Na_2SeO_4 weakens memory formation of CTAs presumably
335 through activation of PP2A.

4.1 CTA Induction

Due to the fact that the learning and memory paradigm of conditioned taste aversion is sensitive to the effects of toxic substances, it was imperative to understand the dose-response relationships of Na_2SeO_4 toxicity before studying its effects on learning and memory. Since there is no literature regarding what constitutes a dose of Na_2SeO_4 that will modulate PP2A activity without showing toxic effects; experimentation was required to determine what constitutes such a dose. Previous studies addressing selenium salt dose toxicity have used both in vivo and in vitro methods (Xiao et al., 2006; Nogueira and Rocha, 2011; Lee and Jeong, 2012).

We are interested in finding a dose that will not confound the behavioral data of a CTA paradigm rather than characterizing the toxic effects of Na_2SeO_4 at the molecular and anatomical level. Therefore, determining which doses of Na_2SeO_4 can serve as a US when paired with a tastant is arguably the most valid method of investigating Na_2SeO_4 toxicity, translationally speaking. After pairing variable doses of Na_2SeO_4 with saccharin (the CS), we determined that a systemically delivered low dose (0.5mg/kg dose) of Na_2SeO_4 is not recognized as a toxin by the test rats. This was concluded with a behavioral assay. A physiological assay of the effects of Na_2SeO_4 could provide useful data as well. That was the objective of our second experiment.

When testing the toxicity of Na_2SeO_4 , we used an acute dose, rather than a chronic treatment of the drug. Like acute doses of Na_2SeO_4 , it has been found that chronic overexposure to selenium salts has additive effects that can enhance the potential risk of toxicity in a dose that would otherwise exhibit benign effects if delivered acutely. Chronic overexposure has been shown to increase the risk of diabetes type II, several types of cancer, and amyotrophic lateral sclerosis (Vinceti et al. 1996, 1998, 2001, 2009, 2010; Stranges et al. 2010). It was concluded that the test animals' exposure to any dose of selenium exceeding their dietary requirement should be as brief as possible to reduce toxic side-effects.

Though the method used to discover toxic doses of Na_2SeO_4 in experiment 1 was translationally valid, this method may lack the specificity that other assays of toxicity have. Physiological assays have the benefit of showing specifically what doses result in damage to tissues and polymers

inside of the cells of affected organisms and the extent of this damage. Additionally, the treatment doses used in experiment 1 were fairly arbitrary. It may be beneficial to characterize the toxic effects of various doses of Na_2SeO_4 closer to the 0.5 mg/kg dose used in experiment 3 in order to see if there are more appropriate doses for various experimental purposes. Furthermore, it may be possible to bypass the toxic signaling of the visceral neuraxis altogether via intracranial cannulation and administration of site specific injections of Na_2SeO_4 to various nuclei (i.e. the amygdala), and thereby reduce the need to characterize the toxicity of various systemic doses. Despite these limitations, the results of experiment 1 show that a systemic dose of 0.5 mg/kg Na_2SeO_4 does not induce a CTA when paired with the CS; therefore it was concluded to be a suitable dose to test the effects of Na_2SeO_4 injections on the learning and memory processes of CTA in experiment 3.

4.2 c-Fos Induction

In experiment 2, we looked at the effects that systemic injections of Na_2SeO_4 had on the activation of a series of brain regions, including the visceral neuraxis. These regions included the parabrachial nucleus (PBN) and the nucleus of the solitary tract (NTS) of the hindbrain and the central amygdala (CeA), the paraventricular nucleus (PVN), and the supraoptic nucleus (SON) of the forebrain. Neural activation was determined by quantifying the immunoreactivity of the inducible transcription factor c-Fos, an antigen commonly used as marker of neural activity. Though there was not a consistent overall pattern of post treatment c-Fos induction, some patterns were apparent when the nuclei were grouped by functional circuitry. The c-Fos induction in the brainstem (PBN and NTS) showed a pattern of neural activation that most resembled a dose-response effect. A graded response in these brain areas would not be surprising considering that the NTS and PBN serve as the first and second sensory relays, respectively, for both taste and visceral signals coming into the brainstem and likely involve relatively little signal processing. Specifically, this dose dependent neural activation in the visceral neuraxis shows that the low dose of Na_2SeO_4 activates these nuclei similarly to that of vehicle injections; while the higher doses treatments results in activity comparable to that of LiCl. This is consistent with the behavioral evidence of experiment 1 showing that a low dose of Na_2SeO_4 does not cause a CTA, but higher doses do in a graded fashion. It is unknown whether Na_2SeO_4 is transduced as a toxin via the vagal pathway or the area postrema because c-Fos induction is not a reliable marker of neural activation in the area postrema.

392 The c-Fos induction patterns in the CeA showed more variability than any other brain region
393 observed. There was an unusually high count of c-Fos in the negative control group as well as a lack
394 of a significant difference between the negative control group and the positive control group in this
395 experiment. This led to conflicting interpretations of the results in this brain region. There may
396 be a dose-response effect, like that in the hindbrain, or possibly an all or nothing effect in which
397 the 1 mg/kg of Na₂SeO₄ is the only dose that induced significantly higher c-Fos than the negative
398 controls. Evidence shows that the CeA plays an important role in associative processes important for
399 the learning and memory of a CTA. It is possible that the low dose does not generate a strong enough
400 toxic signal to the CeA to result in significant neural activation in this brain region. Alternatively,
401 activation of PP2A in the CeA, via Na₂SeO₄ administration, may have blocked learning and memory
402 processes via some molecular pathway that also induces c-Fos expression.

403 Hypothalamic areas (the SON and the PVN) were sampled due to their involvement in
404 stress responses. These areas show activation in response to LiCl injections as well as many other
405 stimuli that can serve as an US. We observed little or no c-Fos induction when the rats were injected
406 with Na₂SeO₄. These results were unusual and could mean that Na₂SeO₄ does not induce a stress
407 response in animals after treatment, even at doses that induce conditioned taste aversions. Another
408 possibility is that Na₂SeO₄ (i.e. activated PP2A) is interfering with a metabolic pathway which is
409 necessary for c-Fos expression. These two possibilities may not be mutually exclusive, given that
410 the interference of some aspect of the c-Fos pathway in the PVN and NTS could hypothetically be
411 responsible for the diminished stress response that we observed. It would be interesting to see if
412 sodium selenate's failure to activate the PVN and NTS was due to an insufficient dose size. It is
413 possible that a much higher dose of Na₂SeO₄ will result in an increase in c-Fos expression in the PVN
414 and NTS. c-Fos was used because it is a known marker of neural activity; but there are several caveats
415 to c-Fos studies. c-Fos is not a marker for neural activity in all types of neurons (Morgan et al., 1987).
416 Other methods of marking neural activity may be appropriate as well, if not more appropriate; such
417 as quantifying pERK and zif-268 mRNA (Yong-Jing and Ru-Rong, 2009; Zhang et al. 1995). The
418 interval of time between administration of each treatment and the euthanization of the test animals
419 was chosen because it was previously shown that, 2 hours post Na₂SeO₄ administration, certain
420 serine/threonine substrates will be dephosphorylated (Corcoran et al., 2010). Though it has been
421 shown that Na₂SeO₄ crosses the blood brain barrier, we used a systemic administrative route in this
422 experiment, so it is not conclusive whether neural activation was due to peripheral activation of the

visceral neuraxis or due to direct intracellular effects on central neurons. It would be interesting to see how other routes of administration affect c-Fos induction.

The results of experiment 2 were consistent with the results of experiment 1, in that the visceral neuraxis was activated in a dose dependent manner similar to the induction of a CTA. Additionally, we observed no activation of the visceral neuraxis or stress response associated hypothalamic areas by the 0.5 mg/kg dose of Na_2SeO_4 . This confirms that the low dose of Na_2SeO_4 is likely a non-toxic dose that did not exacerbate the effects of the US in experiment 3.

4.3 CTA Attenuation

The ultimate goal of this study was to explore the presumed role PP2A plays in the processes involved in learning and memory via systemic injections of sodium selenate. After a non-toxic dose of Na_2SeO_4 was determined, we used this dose to look at the effects of Na_2SeO_4 on conditioned taste aversion learning and memory retention. We used the low dose (0.5 mg/kg) of Na_2SeO_4 as a pretreatment before pairing saccharin (the CS) with LiCl (the US). The results showed that the pretreatment of Na_2SeO_4 reduced the magnitude of the conditioned taste aversion and also caused the CTA to extinguish very rapidly. Due to sodium selenate's known specificity and agonist effects on PP2A, we hypothesized that this was likely due to the activation of PP2A and ultimately an increased removal of phosphate groups from signaling protein substrates. Due to evidence that the manipulation of certain kinases and phosphatases modulates the strength of learned associations and retention of conditioned taste aversions (there is evidence that this occurs in other learning and memory paradigms as well) we concluded that the results of experiment 3 were likely due to effects on associative processes or consolidation via alterations in intracellular signaling and gene expression. However, we cannot rule out the possibility that these experimental effects may have been due to manipulations of the taste and toxic transduction pathways.

The dose of sodium selenate used in experiment 3 was chosen because it did not induce a CTA in experiment 1 and was deemed non-toxic. The timing of the pre-treatment administration was chosen in order to further reduce the chance of strengthening the toxic signal of the US (CTAs are resistant to backwards conditioning) and because past literature has shown that Na_2SeO_4 has

450 effects on substrate phosphorylation 2 hours after an injection (Corcoran et al. 2010). Systemic
451 injections were deemed appropriate for this study due to sodium selenate's ability to penetrate the
452 blood brain barrier.

453 It would be interesting to give direct injections of Na_2SeO_4 to the CeA, or other nuclei of
454 interest, via a cannula (rather than a systemic injection). This would help determine if there are
455 non-specific effects of Na_2SeO_4 on peripheral organs that are confounding the data.

456 Additionally, it is possible that sodium selenate's effects were not due to activation of PP2A.
457 Though other evidence would suggest that this is likely the mechanism through which Na_2SeO_4
458 caused our results, a phosphatase assay should be conducted to verify the efficacy of Na_2SeO_4 .

459 Our results have important contributions to basic research and practical implications as well.
460 Research regarding processes involved in the negative regulation of learning and memory has been
461 neglected in the past; yet negative regulation is likely to be just as critical as positive regulation for
462 the normal function of learning and memory processes. To date, little is known about the role of
463 PP2A in learning and memory processes and even less is known about its involvement in CTA. For
464 practical purposes, knowledge of the effects of Na_2SeO_4 on learning and memory could be important
465 information due to certain possible medical applications. Na_2SeO_4 has been implicated as a possible
466 treatment for Alzheimer's disease and cancer. If it is ever deemed appropriate for clinical use, it
467 would be important to know how this treatment effects memory as a side effect. Likewise, selenium
468 is essential in a normal diet. An understanding of how non-toxic levels of it effect cognitive processes
469 is of some value. Additionally, due to the fact that Na_2SeO_4 can be used supplementally (if the
470 dose is carefully monitored), it may be worthwhile to test it as a possible treatment for maladaptive
471 forms of memory such as post-traumatic stress disorder.

⁴⁷² **4.4 selenate rescue of CTA in tau mice**

⁴⁷³ **4.5 selenate inhibition of phospho-MAP kinase, but not c-Fos**

⁴⁷⁴ Chen et al PLOSone 2014 Phosphoproteomic profiling – check to see if anything interesting about

⁴⁷⁵ MAPK

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482 **5.2 Disclosures**

483 The authors have no conflicts of interest.

484 6 Figure Legends

485 **Figure 1. Initial magnitude of a conditioned taste aversion induced by sodium selenate.**

486 Expression of CTA following injections of variable doses of sodium selenate shown by quantification
487 of first day 2-bottle preference test. Rats received an injection of one of four doses of sodium selenate
488 20 minutes after receiving 10 minutes access to saccharin. The vehicle injected group (0.0 mg/kg
489 sodium selenate dose) showed a high preference for saccharin after the pairing. The 1.0 and 2.0
490 mg/kg sodium selenate dose group show a significantly lower saccharin preference than the vehicle
491 control group, * $p < 0.05$. The 0.5 mg/kg sodium selenate dose group showed a significantly higher
492 saccharin preference than the 2.0 mg/kg sodium selenate dose group, ? $p < 0.05$.

493 **Figure 2. Extinction of a conditioned taste aversion induced by sodium selenate.**

494 Expression of CTA following injections of variable doses of sodium selenate shown by quantification
495 of 14 days of 2-bottle preference test. Rats received an injection of one of four doses of sodium
496 selenate 20 minutes after receiving 10 minutes access to saccharin. The vehicle injected group (0.0
497 mg/kg sodium selenate dose) showed a high preference for saccharin after the pairing. The 1.0
498 and 2.0 mg/kg sodium selenate dose group show a significantly lower saccharin preference than the
499 vehicle control group that extinguishes before the end of the 14 days of preference testing, * $p < 0.05$.
500 The 0.5 mg/kg sodium selenate dose group did not show a significantly lower saccharin preference
501 than the vehicle control group.

502 **Figure 3. Example photomicrographs of c-Fos induction by sodium selenate.**

503 the nucleus of the solitary tract (A), parabrachial (B), central amygdala (C), paraventricular nucleus
504 (D), and supraoptic nucleus (E).

505 [Insert example photomicrographs here]

506 Figure 3A NTS.

507 Figure 3B PBN.

508 Figure 3C CEA.

509 Figure 3D PVN.

510 Figure 3E SON.

511 **Figure 4. Quantification of c-Fos induction.** c-Fos positive immunoreactivity 2 hours
512 after an injection of variable doses of sodium selenate or LiCl in the nucleus of the solitary tract (A),
513 parabrachial (B), central amygdala (C), paraventricular nucleus (D), and supraoptic nucleus (E). *
514 $p < 0.05$ vs. 0 mg/kg; ? $p < 0.05$ vs. LiCl-induced c-Fos.

515 **Figure 5. Conditioned taste aversion after sodium selenate pretreatment.** A.
516 Initial magnitude of conditioned taste aversion with sodium selenate pretreatment. Saccharin pref-
517 erence on the first day of 2-bottle extinction testing. NaCl or selenate pre-treatment alone did not
518 induce CTA. LiCl induced a significant CTA in NaCl-pretreated rats but not in selenate pretreated
519 rats. * $p < 0.05$ vs. NaCl-NaCl group

520 B. Extinction of conditioned taste aversion with sodium selenate pretreatment. Expression of CTA
521 following pretreatment of either vehicle or sodium selenate paired with vehicle injections or the
522 US shown by quantification of 14 days of 2-bottle preference test. The vehicle-vehicle treatment
523 group showed a high preference for saccharin after the pairing. The selenate pretreatment
524 group that received a vehicle injection showed no significant difference from the vehicle-vehicle
525 treatment group. The vehicle-LiCl treatment group showed a CTA throughout all 14 days of
526 the 2-bottle preference test, * $p < 0.05$. The selenate-LiCl treatment group only showed a CTA
527 on day 1 of the 14 days of 2-bottle preference test that immediately extinguished, * $p < 0.05$.

7 References

1. Runyan JD, Moore AN, Dash PK. 2005. A role for prefrontal calcium-sensitive protein phosphatase and kinase activities in working memory. *Learn Mem.* 12:
2. Mansuy IM, Shenolikar S. 2006. Protein serine/threonine phosphatases in neuronal plasticity and disorders of learning and memory. *Trends Neurosci.* 29:
3. Yamashita T, Inui S, Maeda K, Hua DR, Takagi K, et al. 2006. Regulation of caMKII by alpha4/PP2Ac contributes to learning and memory. *Brain Res.* 1082:
4. Munton RP, Vizi S, Mansuy IM. 2004. The role of protein phosphatase-1 in the modulation of synaptic and structural plasticity. *FEBS Lett.* 567:
5. Haege S, Galetzka D, Zechner U, Haaf T, Gamberdinger M, et al. 2010. Spatial learning and expression patterns of pP1 mRNA in mouse hippocampus. *Neuropsychobiology.* 61:
6. Cohen P. 1989. The structure and regulation of protein phosphatases. *Annu Rev Biochem.* 58:
7. Corcoran NM, Martin D, Hutter-Paier B, Windisch M, Nguyen T, et al. 2010. Sodium selenate specifically activates pP2A phosphatase, dephosphorylates tau and reverses memory deficits in an alzheimer's disease model. *J Clin Neurosci.* 17:
8. van Eersel J, Ke YD, Liu X, Delerue F, Kril JJ, et al. 2010. Sodium selenate mitigates tau pathology, neurodegeneration, and functional deficits in alzheimer's disease models. *Proc Natl Acad Sci USA.* 107:
9. Shultz SR, Wright DK, Zheng P, Stuchbery R, Liu S-J, et al. 2015. Sodium selenate reduces hyperphosphorylated tau and improves outcomes after traumatic brain injury. *Brain.* 138:
10. Jones NC, Nguyen T, Corcoran NM, Velakoulis D, Chen T, et al. 2012. Targeting hyperphosphorylated tau with sodium selenate suppresses seizures in rodent models. *Neurobiol. Dis.* 45:
11. Ahmed T, Blum D, Burnouf S, Demeyer D, Buee-Scherrer V, et al. 2015. Rescue of impaired late-phase long-term depression in a tau transgenic mouse model. *Neurobiol Aging.* 36:
12. Koller L, Exon J. 1986. The two faces of selenium-deficiency and toxicity—are similar in animals

- and man. *Can. J. Vet. Res.* 50:
13. Lee KH, Jeong D. 2012. Bimodal actions of selenium essential for antioxidant and toxic pro-oxidant activities: The selenium paradox (review). *Mol Med Rep.* 5:
 14. Kryukov G, Castellano S, Novoselov S, Lobanov A, Zehtab O, et al. 2003. Characterization of mammalian selenoproteomes. *Science.* 300:
 15. Spallholz J, Hoffman D. 2002. Selenium toxicity: Cause and effects in aquatic birds. *Aquat. Toxicol.* 57:
 16. Hurd-Karrer AM, Poos FW. 1936. Toxicity of selenium-containing plants to aphids. *Science.* 84:
 17. Hanson B, Lindblom SD, Loeffler ML, Pilon-Smits EAH. 2004. Selenium protects plants from phloem-feeding aphids due to both deterrence and toxicity. *New Phytol.* 162:
 18. Freeman J, Lindblom S, Quinn C, Fakra S, Marcus M, Pilon-Smits E. 2007. Selenium accumulation protects plants from herbivory by orthoptera via toxicity and deterrence. *New Phytol.* 175:
 19. Vickerman D, Trumble J. 1999. Feeding preferences of *Spodoptera exigua* in response to form and concentration of selenium. *Arch. Insect Biochem. Physiol.* 42:
 20. Heinz GH, SANDERSON CJ. 1990. Avoidance of selenium-treated food by mallards. *Environmental Toxicology and Chemistry.* 9:
 21. Wiemeyer SN, Hoffman DJ. 1996. Reproduction in eastern screech-owls fed selenium. *The Journal of Wildlife Management.* 60:
 22. Freeman J, Quinn C, Lindblom S, Klamper E, Pilon-Smits E. 2009. Selenium protects the hyperaccumulator *Stanleya pinnata* against black-tailed prairie dog herbivory in native seleniferous habitats. *Am. J. Bot.* 96:
 23. Pfister JA, Gardner DR, Cheney CC, Panter KE, Hall JO. 2010. The capability of several toxic plants to condition taste aversions in sheep. *Small Ruminant Research.* 90:
 24. Stowe HD, Eavey AJ, Granger L, Halstead S, Yamini B. 1992. Selenium toxicosis in feeder pigs. *J Am Vet Med Assoc.* 201:
 25. Hladun KR, Smith BH, Mustard JA, Morton RR, Trumble JT. 2012. Selenium toxicity to honey

- bee (*apis mellifera* l.) pollinators: Effects on behaviors and survival. *PLoS ONE*. 7:
26. De La Riva DG, Vindiola BG, Castaneda TN, Parker DR, Trumble JT. 2014. Impact of selenium on mortality, bioaccumulation and feeding deterrence in the invasive argentine ant, *linepithema humile* (hymenoptera: Formicidae). *Sci. Total Environ.* 481:
 27. Franke KW, Potter VR. 1936. The ability of rats to discriminate between diets of varying degrees of toxicity. *Science*. 83:
 28. Franke KW, Potter VR. 1935. A new toxicant occurring naturally in certain samples of plant foodstuffs IX. toxic effects of orally ingested selenium. *J Nutr.* 10:
 29. Franke KW, Moxon AL. 1937. The toxicity of orally ingested arsenic, selenium, tellurium, vanadium and molybdenum. *J Pharmacol Exp Ther.* 61:
 30. Smith MI, Stohlman EF, Lillie RD. 1937. The toxicity and pathology of selenium. *J Pharmacol Exp Ther.* 60:
 31. Schroeder H. 1967. Effects of selenate, selenite and tellurite on the growth and early survival of mice and rats. *J. Nutr.* 92:
 32. Shearer T, Ridlington J. 1976. Fluoride-selenium interaction in the hard and soft tissues of the rat. *J. Nutr.* 106:
 33. Paul M, Mason R, Edwards R. 1989. Effect of potential antidotes on the acute toxicity, tissue disposition and elimination of selenium in rats. *Res. Commun. Chem. Pathol. Pharmacol.* 66:
 34. Provenza FD, Kimball BA, Villalba JJ. 2000. Roles of odor, taste, and toxicity in the food preferences of lambs: Implications for mimicry in plants. *Oikos*. 88:
 35. Corcoran NM, Hovens CM, Michael M, Rosenthal MA, Costello AJ. 2010. Open-label, phase i dose-escalation study of sodium selenate, a novel activator of pP2A, in patients with castration-resistant prostate cancer. *Br J Cancer*. 103:

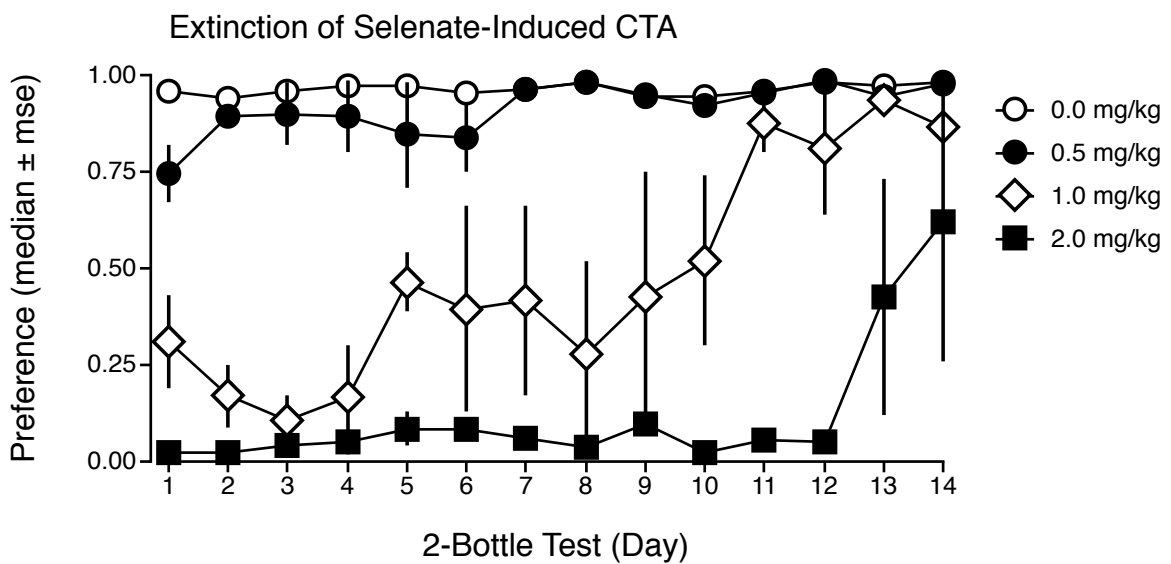
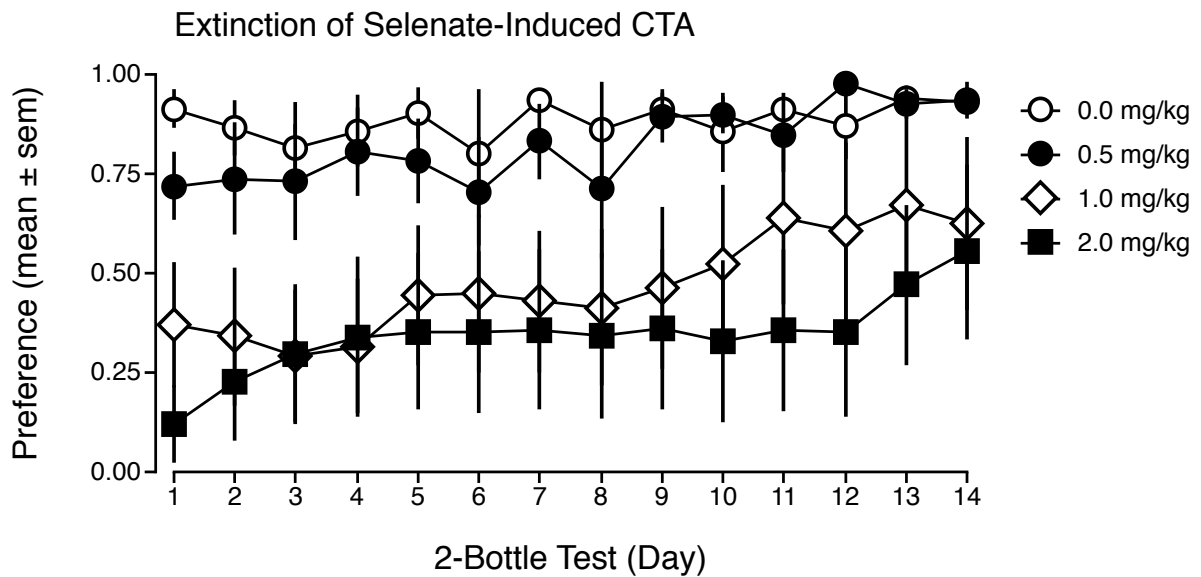
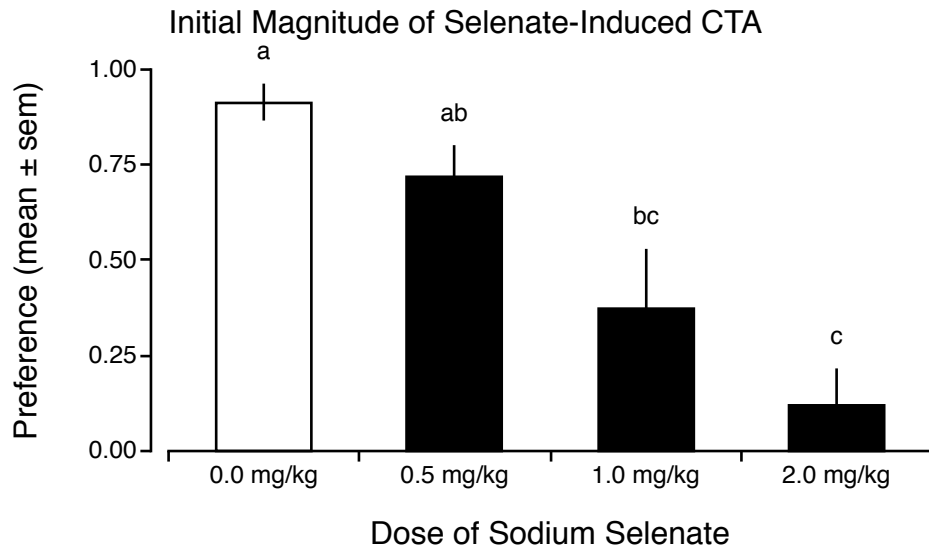


Figure 1

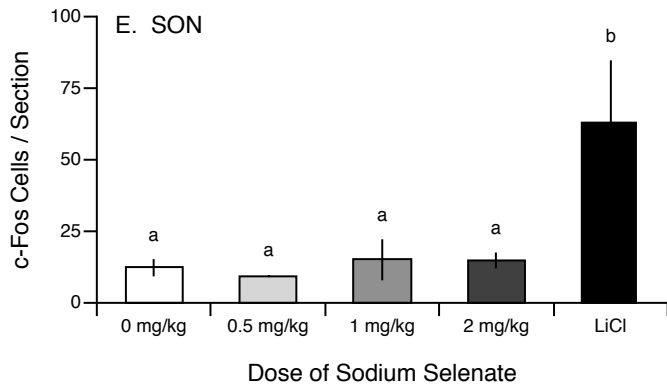
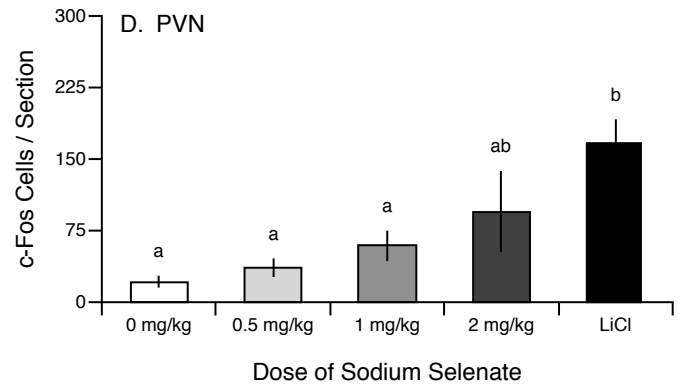
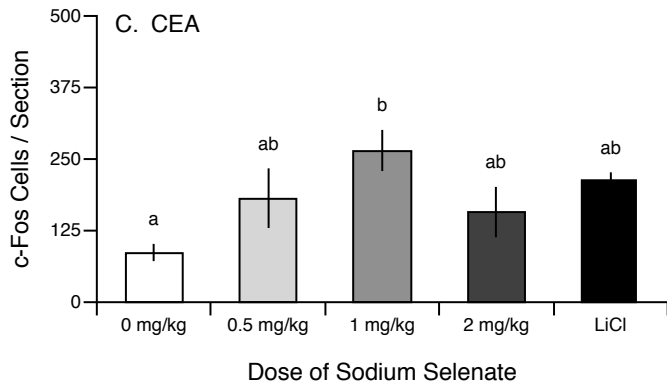
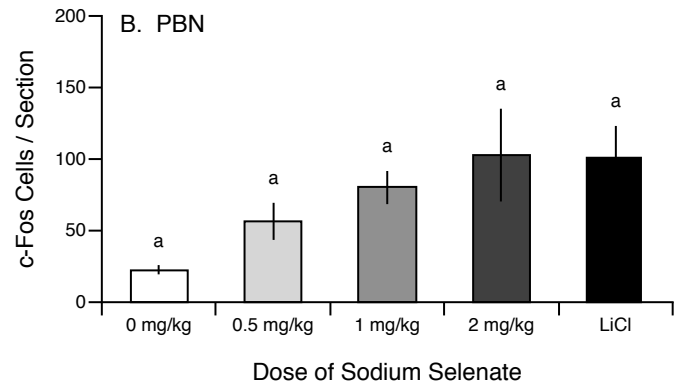
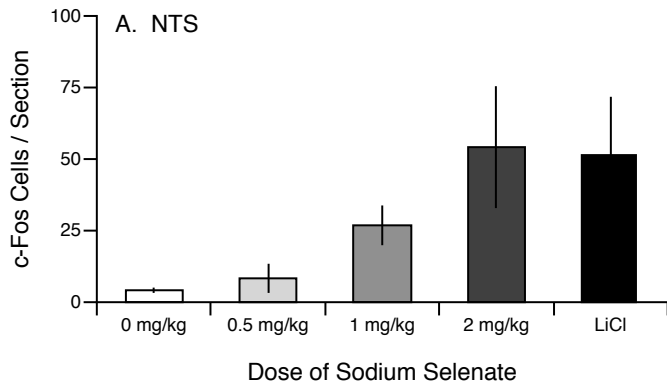
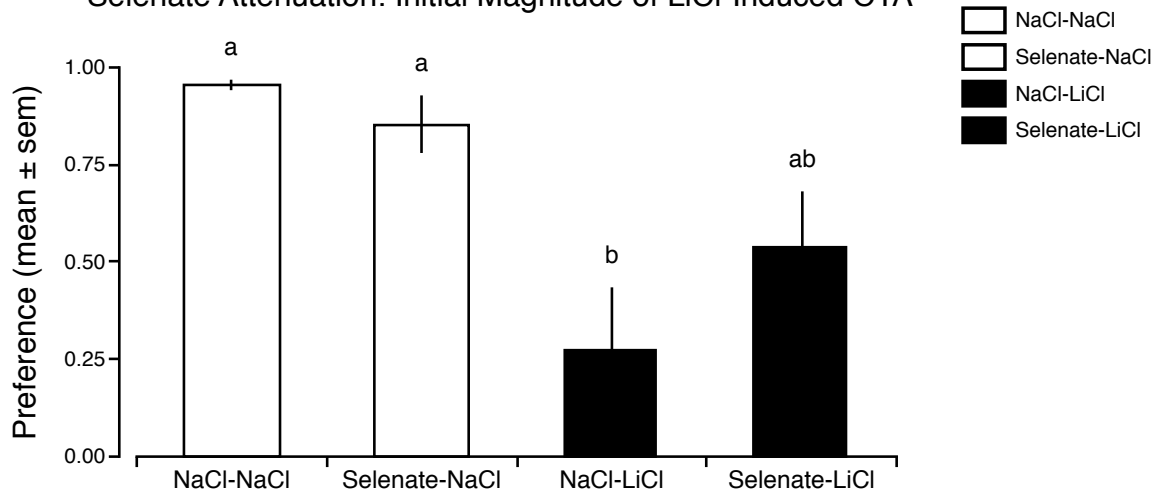
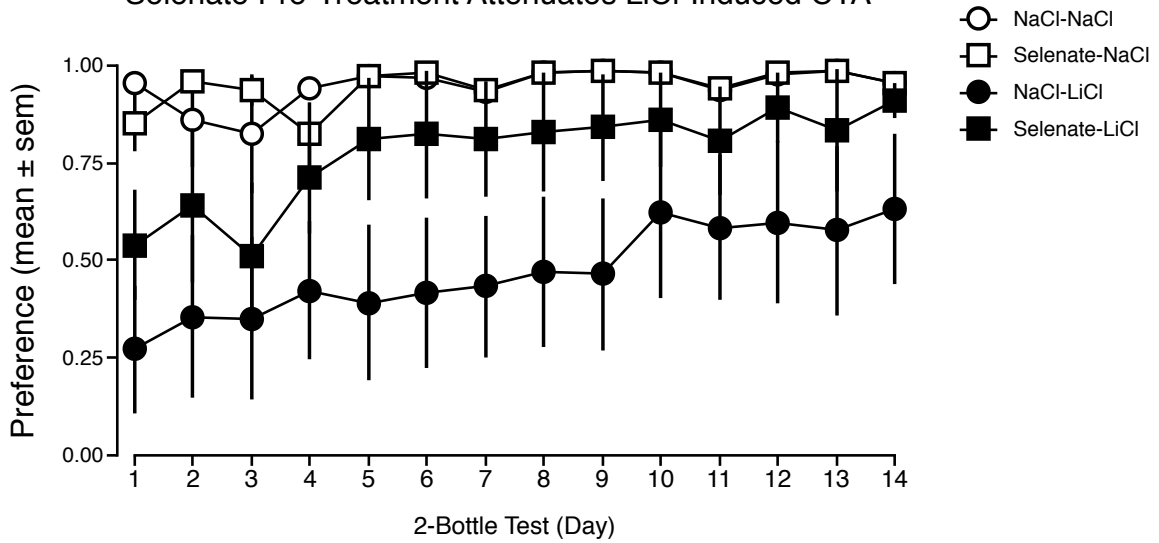


Figure 2

Selenate Attenuation: Initial Magnitude of LiCl-Induced CTA



Selenate Pre-Treatment Attenuates LiCl-Induced CTA



Selenate Pre-Treatment Attenuates LiCl-Induced CTA

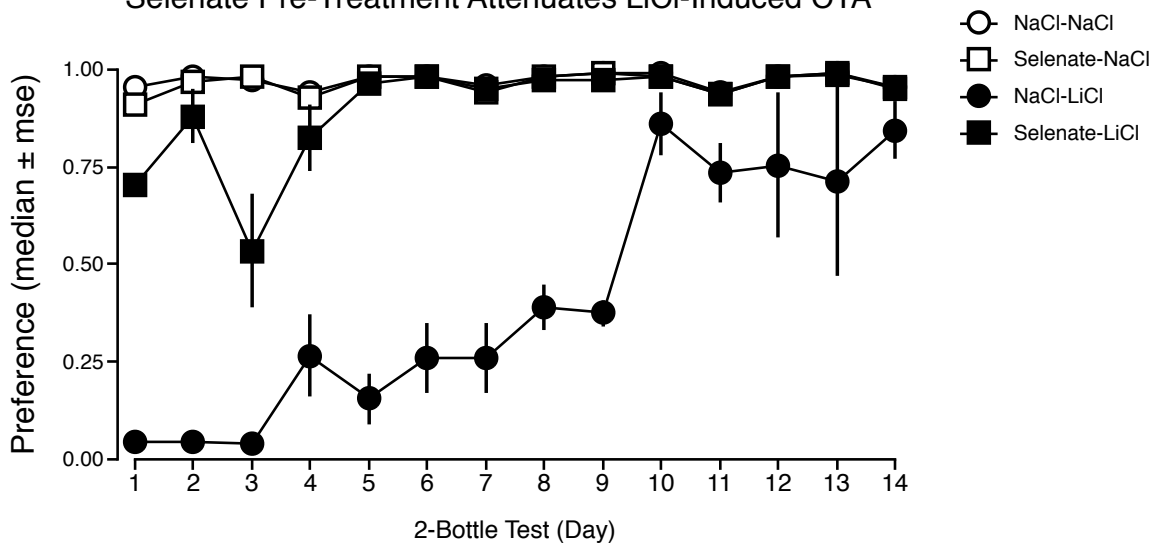


Figure 3