

Molecular Techniques

- Disclaimer
- Nucleic Acids
- Proteins

Haupt, CMN, 9-30-11

3 Goals in Molecular Biology

Identify

All nucleic acids (and proteins) are chemically identical in aggregate - need to identify individual species

Amplify

The amount of an individual gene, mRNA species, or protein is vanishingly small

Visualize

Nucleic acids and proteins are invisible, so need to stain or label to detect and localize.

Nucleic Acids: DNA and RNA

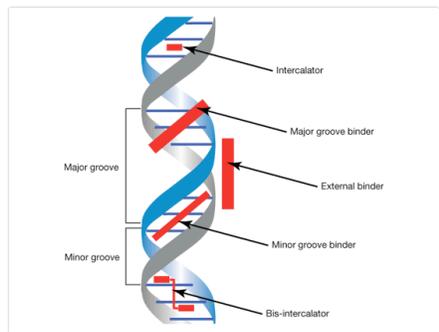
- localization
 - usually nucleus (DNA) or peri-nuclear (mRNA)
- amount
- length (size)
 - in base pairs (bp)
- charge
 - always negative
- sequence
 - A,T(U) ,C,G
- epigenetic modifications
 - proteins bound to DNA or RNA
 - Modification of nucleotide bases, e.g. methylation of DNA

Localization of Nucleic Acids

- **Generic Nucleic Acid Stains**

DAPI for DNA in nucleus

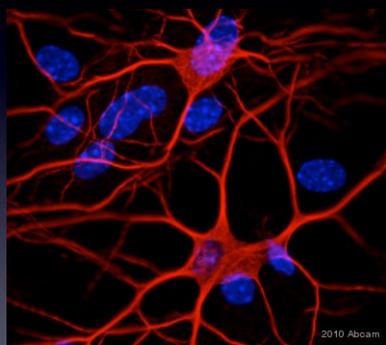
Ethidium Bromide, Syber Green in gels



<http://www.invitrogen.com/site/us/en/home/References/Molecular-Probes-The-Handbook/Nucleic-Acid-Detection-and-Genomics-Technology/Nucleic-Acid-Stains.html>

Example of DNA stain

DAPI stain of DNA in nucleus of cell



Blue = DAPI, DNA in nucleus
Red = MAP2, neuronal cytoplasm

2010 Abcam

<http://www.abcam.com/index.html?pageconfig=reviews&intAbreviewID=21057&intAbID=5392>

Localization of Nucleic Acids

- **In Situ Hybridization**

RNA is single-stranded, but can form a double-helix with a complementary strand

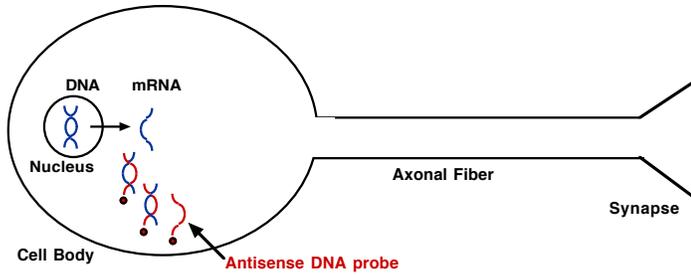
Stick a labeled complementary stretch of DNA or RNA to the mRNA within a tissue section

For example:

DNA probe: ATCCGCATTAG
RNA in blot: TTAGCTTTAGGAGTAATCCGAATATGGC

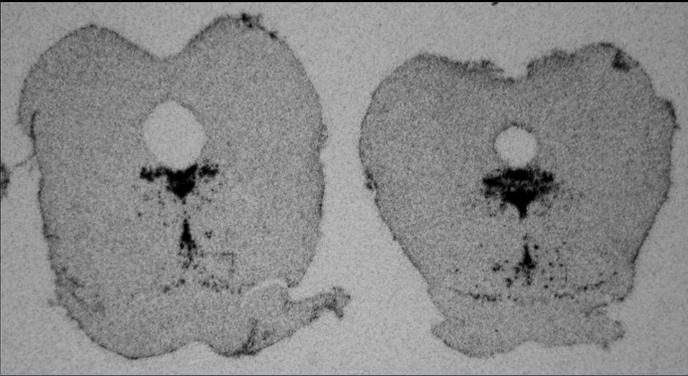
every T in probe is radioactive

use **in situ hybridization** to detect mRNA for enzymes, transporters, or neuropeptides



Label DNA or RNA probe with radioactive nucleotides, or fluorescent nucleotides or with a chemical that can be detected with antibodies
Specific to sequence of target mRNA
Doesn't distinguish different sizes of mRNA species, e.g. alternative splicings of same gene

Example of ISH

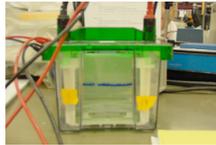
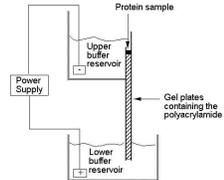
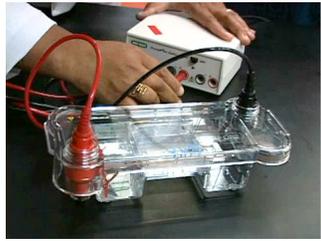


Cell bodies that express mRNA for serotonin transporter

Size Fractionation

- Separation of macromolecules based on size, as measured by rate of travel through gels
- Charged macromolecules impelled to travel through gels by applying an electrical field
- Identify macromolecules by **staining** (e.g. for generic nucleic acids or proteins) or by **probing** (e.g. with specific DNA or antibody probe).

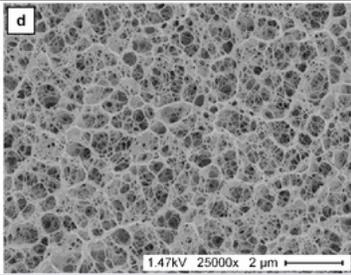
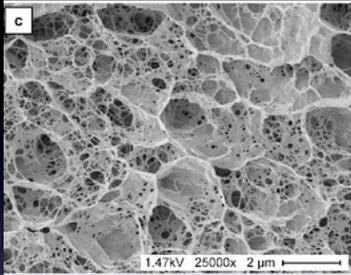
Gel Electrophoresis



<http://newarkbioweb.rutgers.edu/bio301s/lab5-mol-wc-sds-page.htm>

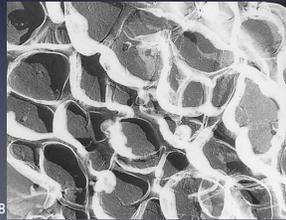
http://web.mit.edu/7.02/virtual_lab/PBC/PBC4Avirtuallab.html

Agarose Gel



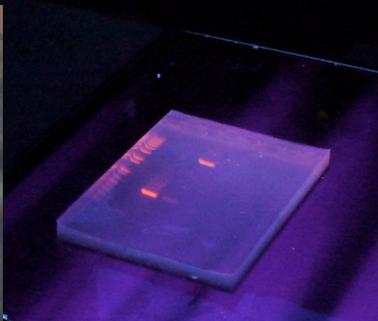
Tuvikene et al., J. Appl. Phycol. 20 (2008)

Polyacrylamide Gel



<http://cellbiologyolm.stevengallik.org/node/78>

Nucleic Acid Stain of Agarose Gel

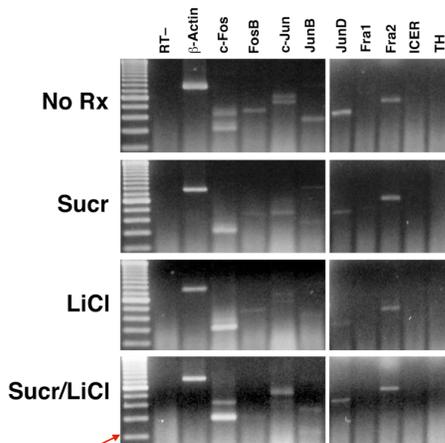


Wikimedia Commons: Agarosegel.jpg

Wikimedia Commons: AgarosegelUV.jpg

Example of DNA in agarose gel

PCR products



ladder = pieces of DNA of known size

Blots

Southern Blot

Separate DNA fragments by size on a gel, then transfer to a nylon membrane (invented by Prof. Southern)

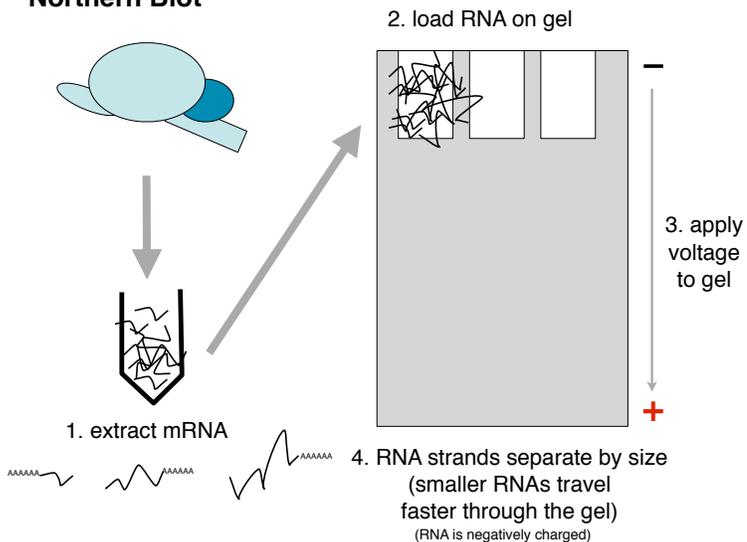
Northern Blot

Separate RNA species by size on a gel, then transfer to a nylon membrane

Western Blot

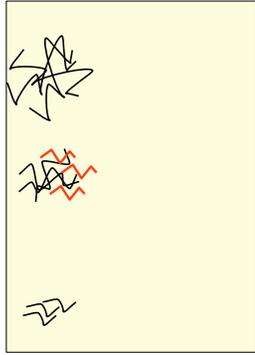
Separate proteins by size on a gel, then transfer to a nylon membrane

Northern Blot



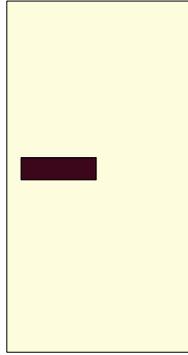
Northern Blot

5. Blot RNA onto nylon membrane



6. Incubate RNA blot with radiolabeled DNA probes

Expose Blot to Film



For example:

DNA probe: **A**TCCGCA**T**TAG (every T is radioactive)
RNA in blot: TTAGCTTTAGGAGTAATCCGAATATGGC

Proteins

- **localization**
depending on function, proteins found throughout cell
- **amount**
- **length (size)**
molecular weight (kiloDaltons; kDa)
- **hydrophobicity**
hydrophilic (water soluble), so likely to be in cytoplasm
hydrophobic (lipid soluble) so likely to be in membrane
- **charge**
variable
- **sequence**
amino acid sequence
- **epitopes**
structural features that may be shared by multiple proteins
recognized by immune system, so can make antibodies
- **postranslational modifications**
phosphorylation

Protein Detection

Protein Stain

Immunohistochemistry

Western blot of protein

Western blot for phosphorylated protein

2-D Gel for detection of all proteins

Example of Protein Stain

Coomassie blue

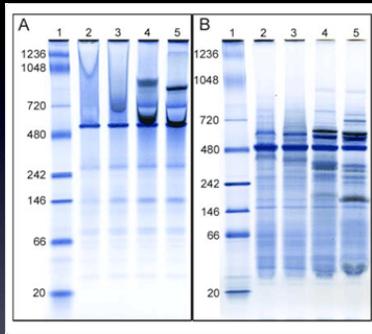
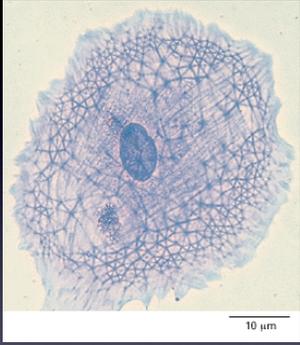


Figure 16-1 from *Molecular Biology of the Cell*
 The cytoskeleton. A cell in culture has been fixed and stained with Coomassie blue, a general stain for proteins. Note the variety of filamentous structures that extend throughout the cell.

Gels were loaded with NativeMark™ standards (lane 1), or 18 μg spinach chloroplast extract. Staining was performed with Colloidal Blue Staining Kit.

<http://www.ncbi.nlm.nih.gov/books/NBK28304/>

Antibodies

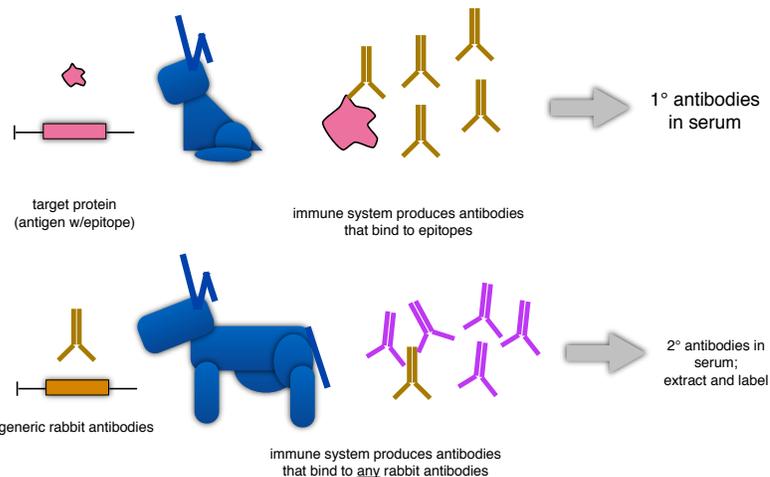
Protein stains do not discriminate different proteins

Protein sequences do not have complements

Immune system produces antibodies that recognizes specific structural features on the surface of proteins and other chemicals (epitopes)

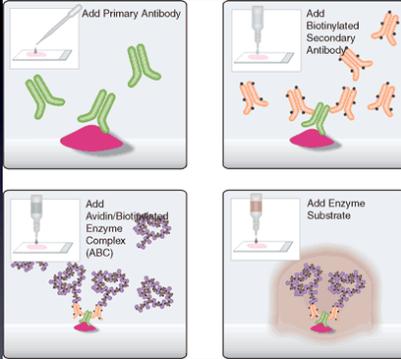
Label antibodies with fluorescent, enzymatic, or radioactive tags

Antibody Production



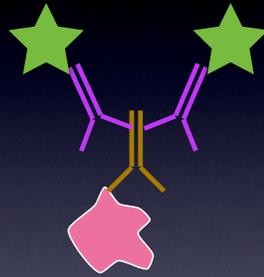
Antibody Detection

Chromogenic



epitope <- 1°Ab <- many 2° Ab <- enzyme complexes <- colored reaction product

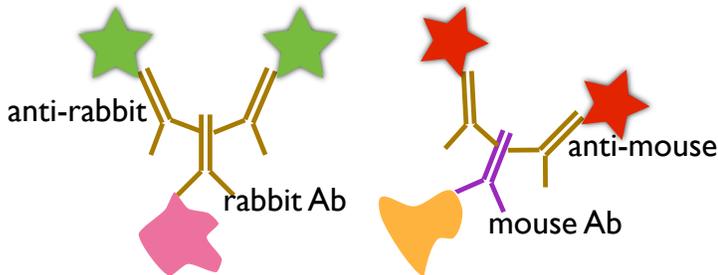
Fluorescent



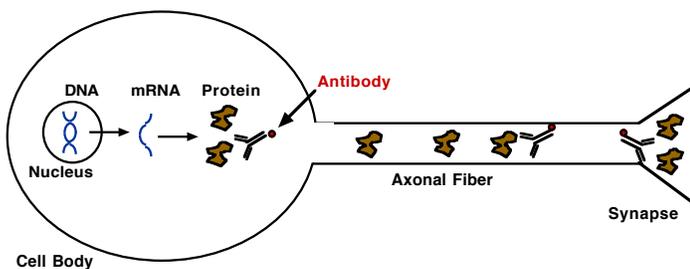
epitope <- 1°Ab <- many labeled 2° Ab <- fluorescence under UV light

www.vector labs

Double-labeling with 2-different species of antibodies



use immunohistochemistry to detect chemicals, proteins



Pros:

- Gives precise anatomical localization
- detects specific molecular feature (epitope) of protein
- can be used with non-proteins (e.g. 5HT, cAMP); anything that immune system can recognize

Cons:

- Antibody may be non-specific; will stick to other proteins with same epitope
- Cannot determine overall protein size or charge

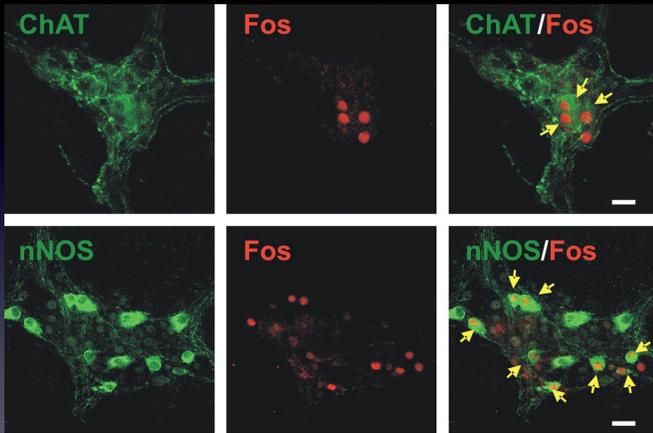
Chromogenic Immunohistochemistry



TH immunostaining of Dopaminergic cells in rat Substantia Nigra

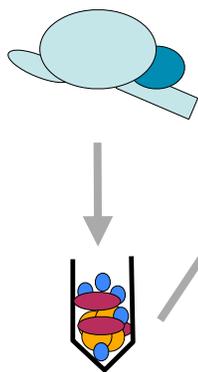
<http://www.psy.herts.ac.uk/res/an-models.html>

Immunofluorescence



Zhou S et al. Am J Physiol Gastrointest Liver Physiol
2008;294:G1201-G1209

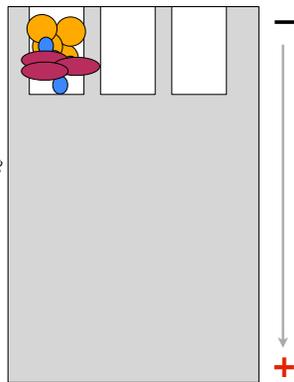
Western Blot



1. extract proteins

SDS denatures &
gives proteins (-) charge

2. load proteins on gel

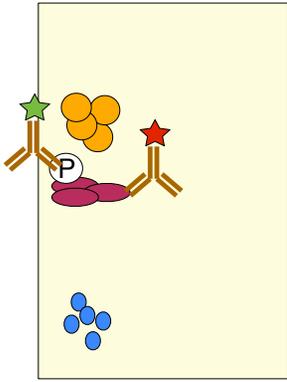


4. proteins separate by size
(smaller proteins travel
faster through the gel)

3. apply voltage
to gel

Western Blot

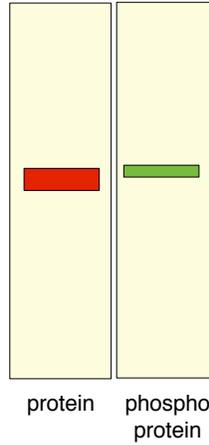
5. Blot proteins onto nylon membrane



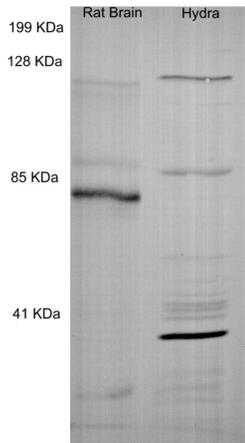
6. Visualize proteins using labeled antibodies

7. Visualize phosphorylated proteins using phospho-specific antibodies

Developed Blot



Example of western blot

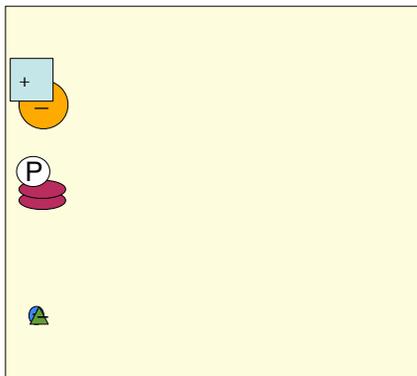


Western blot analysis of NMDA receptor proteins in rat brain (*left lane*) and whole hydra tissue (*right lane*). Numbers indicate the position of Kaleidoscope molecular weight standards. All minor bands were present in the absence of primary antibody. The most prominent band in each lane was only present when primary antibody was included

Immunocytochemical evidence for an NMDA1 receptor subunit in dissociated cells of *Hydra vulgaris*
Scappaticci et al, 2004

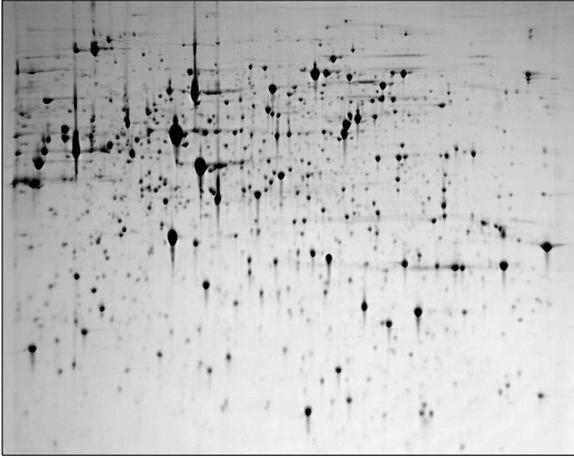
2-D Gel

1. Separate proteins by size on vertical axis



2. Separate proteins by charge on horizontal axis

2-D Gel stained with coomassie blue



<http://abdn.ac.uk/proteome/ap-Saccharomyces-cerevisiae-2d-gel.htm>